

# ACHENE MORPHOLOGY AND PERICARP ANATOMY OF ANEMONE, HEPATICA, AND PULSATILLA (ANEMONINAE, RANUNCULACEAE)

Balkrishna Ghimire<sup>1</sup> D, Dabin Yum<sup>2</sup>, Jae Hyeun Kim<sup>2</sup> and Mi Jin Jeong<sup>2\*</sup>

<sup>1</sup>Division of Forest Biodiversity, Korea National Arboretum, Pocheon 11186, Korea <sup>2</sup>Division of Plant Resources, Korea National Arboretum, Pocheon 11186, Korea

Received June 22, 2020; revision accepted February 16, 2021

The achene morphology and pericarp anatomy of 12 taxa representing three genera (*Anemone*, *Hepatica*, and *Pulsatilla*) of the subtribe Anemoninae were investigated using microtome and light microscopy to evaluate the taxonomic implications of achene characters. The achenes of *Anemone* were elliptical or obvoid and beaked, whereas the achene of *Hepatica* and *Pulsatilla* were obvoid and elliptical, respectively. Noticeable variations in both quantitative and qualitative features of achenes were observed among the species of the three genera. One-way analysis of variance indicated that the quantitative achene variables among the species were highly significant (P<0.001). Pearson's correlation coefficient also showed a significant correlation between different achene variables. The pericarp structure, particularly the number of cell layers and cell forms in the exocarp and endocarp, seems to be very useful for species delimitation in *Anemone*. Unweighted paired group analysis showed the utility of achene features for taxonomic groupings of the species within the studied genera. Although the specien samples represented a limited range of taxa, the achene features and pericarp anatomy provided a reasonable source for the taxonomic treatment of the studied genera within the subtribe.

Keywords: achene, Anemoninae, pericarp, Ranunculaceae, taxonomy

# INTRODUCTION

The Ranunculaceae comprises 2,377 accepted species within 65 genera and is one of the largest basal families within Eudicots (Soltis et al., 2005; Simpson, 2006; Heywood et al., 2007; The Plant List, 2013). The plant is distributed worldwide and exhibits wide variations in morphological characters, especially in its fruit type and floral organization (Emadzade et al., 2010). Several classifications have been proposed based on its morphological characters, molecular data, and combined morphological and molecular dataset (Hutchinson, 1923; Janchen, 1949; Johansson and Jansen, 1993; Tamura, 1995; Jansen et al., 1995; Ro et al., 1997; Wang et al., 2009). Based on the basic chromosome number and carpel and fruit types Tamura (1995) divided the family into three subfamilies: Hellebor-

oideae with four tribes, Ranunculoideae with three tribes, and Isopyroideae with four tribes.

Anemoneae is one of the three tribes of the subfamily Ranunculoideae which is traditionally divided into three subtribes: Kingdoniinae, Anemoninae, and Clematidinae. However, in contemporary classifications, based on morphology and molecular sequence data, the subtribe Kingdoniinae, with only one species, Kingdonia uniflora, has been excluded from Anemoneae, even from Ranunculaceae, and is treated as its own family Kingdoniaceae or included within the family Circaeasteraceae (Ren et al., 2004; Wang et al., 2009; APG IV 2016). In addition, Anemoneae, with the remaining two tribes, is strongly inferred as the monophyletic group in phylogenetic analyses (Jansen et al., 1995; Ro et al., 1997; Wang et al., 2009; Jiang, 2017).

<sup>\*</sup> Corresponding author, email: mjjeong121@korea.kr





The subtribe Anemoninae consists of eight genera: Anemoneclema (Franch.) W. T. Wang, Anemone L., Barneoudia C. Gray, Hepatica Miller, Knowltonia Salisb, Metanemone W. T. Wang, Oreithales Schldl., and Pulsatilla Mill (Wang, 1979; Tamura, 1995). Among them, Anemone is the largest one with more than 150 species whereas Anemoneclema, Metanemone, and Barneoudia are monotypic. Molecular phylogenetic studies revealed that Hepatica, Pulsatilla, and Knowltonia are nested within Anemone (Hoot et al., 1994; Barniske, 2009; Meyer et al., 2010; Hoot et al., 2012). In addition, the phylogenetic relationship and taxonomic status of Anemoneclema in the subtribe has remained unclear and recent molecular studies suggested the transfer of the monotypic genus to Clematidinae from Anemoninae (Zhang et al., 2015; Lehtonen et al., 2016).

Fruit and seed morphologies have provided important phylogenetic sources, which are frequently used to differentiate the taxa in different taxonomic ranks. The surface sculpture of fruits, seeds or a combination of both are particularly used for phylogenetic and/or systematic studies (Fukuhara, 1999; Juan et al., 2000; Xu, 2003; Zhang et al., 2005; Amini et al., 2011; Ghimire et al., 2015, 2016a). There have been several studies that emphasize the taxonomic utility of fruit and seed morphological characteristics of different Ranunuculaceae taxa (Cappelletti and Poldini, 1984; Heiss et al., 2011; Ghimire et al., 2015, 2016b; Hadidchi et al., 2019; Ghimire et al., 2020).

The achene anatomy of Anemone has had a long history and proved to be of great value for the infrageneric classification of the genus (Starodubtsev, 1991; Tamura, 1995; Zeman et al., 2008). Zeman et al. (2008) extensively studied the anatomy of 110 species of Anemone and found as many as 25 achene features with potential diagnostic value. Before that Chaudhary and Trifonova (1988) examined the usefulness of fruit anatomy in the systematic position of 11 Nepalese Anemone. Maciejewska-Rutkowska and Antkowska (2013) demonstrated the importance of achene morphology and anatomy in four Polish Anemone. However, in the literature there is a lack of a comparative study of the achenes of Anemone species with other genera of the subtribe Anemoninae. Jung and Heo (2017) listed a few achene anatomical characters of three Anemone, three Hepatica, and two Pulsatilla species in their study but did not discuss the utility of the features within the Anemoninae.

In the present study, we investigated the achene anatomy of 12 species belonging to three genera of the subtribe Anemoninae. The primary objectives of this study were (1) to demonstrate the achene morphological and anatomical features of the included taxa and (2) to understand the taxonomic usefulness of achene features within and between the genera of the subtribe Anemoninae.

## MATERIALS AND METHODS

#### SPECIMENS

More than 240 achenes from 12 taxa representing three genera (*Anemone*, *Hepatica*, and *Pulsatilla*) of the subtribe Anemoninae originating from the seed bank at the Korea National Arboretum, Pocheon, Korea were investigated. Names of the investigated species with their voucher numbers are shown in Table 1.

TABLE 1. Name of taxa with voucher number.

Taxon	Voucher No.
Anemone koraiensis Nakai	L10335
Anemone raddeana Regel	L2030
Anemone reflexa Steph. Ex. Willd	L8045
Anemone narcissiflora L.	L14911
Anemone narcissiflora subsp. crinita (Juz.) Kitag	L10766
Hepatica asiatica Nakai	L3363
Hepatica insularis Nakai	L9859
Hepatica maxima (Nakai) Nakai	2015UL001
Pulsatilla dahurica (Fisch. Ex DC.) Spreng	L12332
<i>Pulsatilla cernua</i> (Thunb.) Bercht. ex J. Presl	L13107
Pulsatilla koreana Nakai	L12512
Pulsatilla tongkangensis Y. N.Lee & T.C.Lee	L12843

## LIGHT MICROSCOPY

At least three to five achenes of each taxa were considered for microtome sectioning. Microtome sections were prepared using the following proce-

dure. Mature achenes were passed through ethanol series (50, 70, 80, 90, 95 and 100%) for dehydration. After complete dehydration, the achenes were infiltered in alcohol/Technovit combinations (3:1, 1:1, 1:3, and 100% Technovit) and then embedded in Technovit 7100 resin. The embedded materials were cut into serial sections of 4-6 µm thickness using a Leica RM2255 rotary microtome (Leica Microsystems GmbH, Germany) with disposable blades, stuck onto a slide glass, and dried using an electric slide warmer for 12 h. The dried slides were stained with 0.1% Toluidine blue 'O' for 60-90 s. rinsed with water and again dried in slide warmer for more than 6 h to remove water. The stained slides were then mounted with Entellan (Merck Co., Germany) and permanent slides were prepared. The permanent slides were examined under an AXIO Imager A1 light microscope (Carl Zeiss, Germany). Photomicrographs were captured with an AxioCam MRc5 attached camera system, and pericarp measurements were made using AxioVision software for Windows (release 4.7, 2008). Multiple image alignment was performed using Photoshop CS for Windows 2010. None of the image-alteration facilities of Photoshop were used to change the original images captured by the camera.

#### MORPHOMETRY AND DATA ANALYSIS

A total of approximately 240 achenes were used for morphometric measurement. Digital images of whole achenes were captured with a Leica DFC420 C multifocal camera attached to a Leica MZ16 FA microscope (Leica Microsystems, Germany). The length and width of 20 seeds from each taxon were measured using Leica LAS V3.8 software for Windows. The biometric data were analyzed statistically. For each seed variable, one--factor analysis of variance (ANOVA) was used to examine the differences in means among the included species. Pearson's correlation coefficients were used to estimate the relationship among the achene length, width, pericarp and endocarp thickness, diameter cross section, and their relative ratios. All statistical analyses were performed using the SPSS statistical program (IBM SPSS Statistics for Windows Version 20.0., IBM Corp., Armonk, USA). Six quantitative and 11 qualitative characters were categorized and coded with binary and/or multistate coding. Cluster analysis using the unweighted paired group method with arithmetic mean (UPGMA) was also performed to verify achene features and it allowed grouping of the

species by using the statistical program PAST ver. 3.25 (Hammer et al., 2001). Character states and their codes are shown in Supplementary File S1.

## RESULTS

Selected stereomicroscopic images of a single achene and light microscopic images of the pericarp structure are shown in Figures 1–6. A comprehensive description of the morphological features of achenes and anatomical features of the pericarp is presented below.

#### ANEMONE L.

Altogether, five taxa of *Anemone* were investigated in this study. The achenes are elliptical or obovoid, brown to pale yellow in color, and flattened (Figs. 1a–e; Table 2). The largest achene was measured in *Anemone narcissiflora* (7.75±0.82 × 5.58±0.66 mm), followed by *A. narcissiflora* subsp. *crinita* (7.24±0.51 × 4.49±0.63 mm), whereas the smallest achene was measured in *A. raddeana* (2.26±0.18 × 1.18±0.11 mm), followed by *A. reflexa* (2.42±0.24 × 0.84±0.09 mm) (Table 3). Out of the five taxa, three (*A. koraiensis*, *A. raddeana*, and *A. reflexa*) have hair on their achene surface and narrow wings, but the remaining



**Fig. 1.** Achenes of Anemone under stereomicroscope (a) A. koraiensis (b) A. reflexa (c) A. raddeana (d) A. narcissiflora subsp. crinita (e) A. narcissiflora Scale bars: **a**,**b**,**c** = 0.5 mm; **d**,**e** = 1 mm

		www.czasopisma.pan.		N www.journa	ls.pan.pl			
Seed coat	Thick and well represented, exotesta single layered, mesotesta 4-5 layered and endotesta single layered	Either represented by parenchymatous layers or crushed. Exotesta single layered elongated cells, mesotesta 2-3 layers of thin walled cells, endotesta single layered thick walled	Represented by crushed layers	3-4 layers of degenerating	Represented by crushed layers	2-3 layers of degenerating cells	2-3 layers of degenerating cells	2-3 layers of degenerating cells
Endocarp	Thin layer of thick-walled rectangular cells	Thick layer of highly lignified palisade like cells	Thick layer of highly lignified palisade like cells	2 to 3 layers, rarely single layer of highly lignified sclereid cells	Thick layer of highly lignified sclereid cells	2 to 3 layers, rarely single layer of highly lignified palisade cells	2 to 3 layers of highly lignified palisade cells	2 to 4 layers of highly lignified sclereid cells
Mesocarp	Well represented, 7-10 cells thick, isobilateral/elongated/ irregular parenchy- matous cells with wavy walls	Well represented, 3-5 cells thick, elongated paren- chymatous cells with straight or wavy walls	Represented by crushed cells	Well represented, 3-5 cells thick, elongated paren- chymatous cells with straight or wavy walls	Represented by crushed cells	1-2 layers of elongated parenchyma cells.	1-2 layers of elongated parenchyma cells	Well represented, 3-5 cells thick, elongated paren- chymatous cells
Exocarp	Cutinized, rectangular cells	Cutinized, rectangle to tangentially elongated	Cutinized, cell crushed	Cutinized, mostly crushed, if present tangentially elongated	Cutinized, cell crushed	Cutinized, tangentially elongated	Cutinized, tangentially elongated	Cutinized, rectangular cells
Shape in CS	Oval	Oval	Oval	Elliptical	Elliptical	Circular	Circular	Circular
Style	Hooked	Hooked	Hooked	Hooked	Hooked	Slender	Slender	Slender
Wings	Very narrow	Narrow	Narrow	Wide	Wide	Narrow	Very narrow	Narrow
Surface	Hairy	Hairy	Hairy	Glabrous	Glabrous	Hairy	Hairy	Glabrous
Color	Pale yellow to brown	Pale yellow to brown	Brown	Pale yellow to brown	brown	Pale yellow to brown	Pale yellow to brown	Dark brown to black
Shape	Elliptical	Elliptical	Elliptical	Obovoid, flattened	Obovoid, flattened	Obovoid	Obovoid	Obovoid
Taxon	A. koraiensis	A. reflexa	A. raddeana	A. narcissi- flora	A. narcissi- flora subsp. crinita	H. asiatica	H. insularis	H. maxima

TABLE 2. Morphological and anatomical features of achene of Anemoninae.

_	Shape	Color	Surface	Wings	Style	Shape in CS	Exocarp	Mesocarp	Endocarp	Seed coat
								with wavy walls		
urica E	lliptical	Brown	Hairy	Narrow	Slender	Circular	Crushed	Represented by crushed cells	Thick layer of highly lignified sclereid cells	Represented by crushed layers
nua ] e	Narrow Slliptical	Brown to black	Hairy	Narrow	Slender	Circular	Crushed	Represented by crushed cells	Thick layer of highly lignified sclereid cells	Represented by crushed layers
eana ] e	Narrow Siliptical	Brown to black	Hairy	Narrow	Slender	Circular	Crushed	Represented by crushed cells	Thick layer of highly lignified sclereid cells	Represented by crushed layers
gkan- ] s e	Narrow Iliptical	Brown to black	Hairy	Narrow	Slender	Circular	Crushed	Represented by crushed cells	Thick layer of highly lignified sclereid cells	2-3 layers of elongated cells with thickened walls

TABLE 3. Achene characters and their measurements in Anemoninae species (mean and standard deviation). L/W = length and width ratio, P/E = pericarp and endocarp ratio, CSD1 = diameter parallel to cotyledon, CSD2 = diameter perpendicular to cotyledon, D1/D2 = CSD1 and CSD2 ratio.

Taxa	Length (mm)	Width (mm)	L/W	Pericarp (µm)	Endocarp (µm)	P/E	CSD1 (mm)	CSD2 (mm)	D1/D2	www.jo
A. koraiensis	$3.34 \pm 0.22$	$1.74 \pm 18$	$1.93 \pm 0.2$	$206.78 \pm 17.47$	$9.79 \pm 1.99$	$21.75 \pm 3.82$	$1.97\pm0.16$	$1.51 \pm 0.13$	$1.31 \pm 0.06$	ourn
A. reflexa	$2.42 \pm 0.24$	$0.84 \pm 0.09$	$2.9\pm 0.34$	$115.1\pm22.86$	$25.4 \pm 4.01$	$4.57 \pm 0.66$	$1.8 {\pm} 0.07$	$1.17 \pm 0.1$	$1.55 \pm 0.07$	ais.
A. raddeana	$2.26{\pm}0.18$	$1.18\pm0.11$	$1.94 {\pm} 0.2$	$56.65 \pm 13.89$	$25.49\pm 5.25$	$2.27\pm0.55$	$1.75 \pm 0.08$	$1.45 \pm 0.09$	$1.21 \pm 0.06$	pan.
A. narcissiflora	$7.75\pm0.82$	$5.58 \pm 0.66$	$1.4 \pm 0.17$	$149.27 \pm 15.3$	$29.84 \pm 4.62$	$5.11 \pm 0.97$	$3.91 \pm 0.15$	$1.82 \pm 0.06$	$2.15\pm0.03$	.pi
A. narcissiflora subsp. crinita	$7.24 \pm 0.51$	$4.49\pm0.63$	$1.58 \pm 0.24$	$43.53\pm6.03$	$18.29 \pm 2.93$	$2.42 \pm 0.37$	$3.13 \pm 0.11$	$1.44 \pm 0.09$	$2.17\pm0.12$	
H. asiatica	$2.11 \pm 0.23$	$1.32 \pm 0.15$	$1.62 {\pm} 0.21$	$99.04 \pm 8.35$	$64.77\pm5.9$	$1.53\pm0.13$	$2.01 \pm 0.1$	$1.94 \pm 0.04$	$1.04 {\pm} 0.04$	
H. insularis	$2.43 \pm 0.19$	$1.41 \pm 0.11$	$1.73 {\pm} 0.16$	$126.85 \pm 13.57$	$79.83 \pm 12.06$	$1.61 \pm 0.15$	$2.02 \pm 0.18$	$2.89 {\pm} 0.12$	$1.07 \pm 0.06$	
H. maxima	$3.35 \pm 0.32$	$1.9 \pm 0.29$	$1.83 \pm 0.4$	$274.38 {\pm} 27.83$	$80.41 \pm 6.84$	$3.43\pm0.43$	$2.98 \pm 0.23$	$2.86 {\pm} 0.25$	$1.04 \pm 0.1$	
P. dahurica	$2.7 \pm 0.3$	$0.87 \pm 0.08$	$3.12 \pm 0.4$	$37.31 \pm 4.32$	$25.33 \pm 3.88$	$1.48 \pm 0.11$	$0.86 \pm 0.11$	$0.85 \pm 0.11$	$1.02 \pm 0.6$	
P. cernua	$3.6 {\pm} 0.19$	$1.05 \pm 0.07$	$3.42 \pm 0.33$	$32.11 \pm 5.78$	$18.24 \pm 3.41$	$1.78 \pm 0.25$	$0.91 \pm 0.07$	$0.85 \pm 0.13$	$1.12 \pm 0.14$	
P. koreana	$3.2 \pm 0.33$	$1.11 \pm 0.17$	$2.93 \pm 0.38$	$34.55 \pm 9.78$	$18.59 \pm 3.23$	$1.78 \pm 0.43$	$1.05 \pm 0.05$	$1.02 \pm 0.08$	$1.03 \pm 0.04$	
P. tongkangensis	$4.07 \pm 0.24$	$1.17 \pm 0.51$	$3.46 {\pm} 0.42$	$58.63\pm8.199$	$32.84 {\pm} 3.35$	$1.78 \pm 0.07$	$1.07 \pm 0.09$	$1.03\pm0.08$	$1.01 \pm 0.05$	
ANOVA	F=451.9, P<0.001	F=424.64, P<0.001	F=101.07, P<0.001	F=400.55, P<0.001	F=300.14, P<0.001	F=334.31, P<0.001	F=244.34, P<0.001	F=206.1, P<0.001	F=115.18, P<0.001	





two (*A. narcissiflora* subsp. *crinita* and *A. narcissiflora*) have glabrous achene surfaces and wide wings. All the species have a hooked persistent style.

As outlined in the cross-section, ellipsoidal achenes are oval, whereas obovoid achenes are elliptical. The cross-section diameter corresponds with the length and width of achenes. In the cross-section, the thickest pericarp was found in *A. koraiensis* (206.78±17.47  $\mu$ m), followed by *A. narcissiflora* (149.27±15.3  $\mu$ m), and the thinnest pericarp measured was in *A. narcissiflora* subsp. *crinita* (43.53±6.03  $\mu$ m), followed by *A. raddeana* (56.65±13.89  $\mu$ m). Interestingly, *A. koraiensis*, which has the thickest pericarp.

exceptionally bears the thinnest endocarp  $(9.79\pm1.99 \ \mu\text{m})$ . The exocarp in *Anemone* taxa is highly cutinized and the cells are usually crushed except *A. koraiensis* and *A. reflexa*, which have rectangular or tangentially elongated exotesta (Figs. 2a-j). The exocarp layer is followed by a mesocarp with parenchyma cells in *A. koraiensis*, *A. reflexa*, and *A. narcissiflora* (Figs. 2b,d,j); however, this region is represented by a few layers of crushed cells in *A. raddeana* and *A. narcissiflora* subsp. *crinita* (Figs. 2f,h). The thickest endocarp was measured in *A. narcissiflora* (29.84±4.62 \ \mum), followed by *A. raddeana* (25.49±5.25 \ \mum). All the observed taxa have lignified (either palisade-like or sclereid-like) endocarp, regardless of the variation



**Fig. 2.** Cross section of Anemone achenes  $(\mathbf{a}, \mathbf{b})$  A. koraiensis  $(\mathbf{c}, \mathbf{d})$  A. reflexa  $(\mathbf{e}, \mathbf{f})$  A. raddeana  $(\mathbf{g}, \mathbf{h})$  A. narcissiflora subsp. crinita  $(\mathbf{i}, \mathbf{j})$  A. narcissiflora. Scale bars:  $\mathbf{a}, \mathbf{c}, \mathbf{e} = 0.5 \text{ mm}; \mathbf{b}, \mathbf{d}, \mathbf{f}, \mathbf{h}, \mathbf{j} = 0.1 \text{ mm}; \mathbf{g}, \mathbf{i} = 1 \text{ mm}$ . Abbreviations: enc – endocarp, exc – exocarp, msc – mesocarp, sc – seed coat, vb – vascular bundle.

in their thickness (Figs. 2b,d,f,h,j,l). In all the species, the seed coat is feebly established, and either represented by a couple of parenchyma layers or containing degenerating cells.

www.czasopisma.pan.pl

# HEPATICA MILL.

Three species of the genus *Hepatica* are included in this study. Achene morphology such as shape, size, color, and anatomy of the pericarp, endocarp, and seed coat displayed relatively similar qualitative features and quantitative measurements. The achenes are stalked, obovoid, pale yellow to brown or black in color, and pubescent (*Hepatica asiatica*, *H. insularis*) or glabrous (*H. maxima*) in the surface texture (Fig. 3; Table 2). *Hepatica maxima* has comparatively larger achenes  $(3.35\pm0.32 \times$  $1.9\pm0.29$ mm) than *H. asiatica*  $(2.11\pm0.23 \times$  $1.32\pm0.15$ mm) and *H. insularis*  $(2.43\pm0.19 \times$  $1.41\pm0.11$ mm). The lateral wings are small in *Hepatica* and the persistent style develops into a long slender beak (Figs. 3a,b,c).

In the cross-section, *Hepatica* achenes are circular in outline (Figs. 4a,c,e). The cross-sectional diameter is equivalent to the size of an achene: the larger the achene, the bigger the endosperm diameter (Table 3). Likewise, the pericarp is thicker in *H. maxima* (274.38 $\pm$ 27.83 µm) than in



**Fig. 3.** Achenes of *Hepatica* under stereomicroscope (a) *H. asiatica* (b) *H. insularis* (c) *H. maxima.* Scale bars = 1 mm

H. asiatica (99.04 $\pm$ 8.35 µm) and H. insularis  $(126.85 \pm 13.57 \ \mu m)$  but the endocarp is relatively comparable in all species: thus, the ratio of the pericarp and endocarp thickness is relatively higher in H. maxima (Table 3). The exocarp is single--layered, cutinized with either tangentially elongated and rectangular (H. asiatica, H. insularis) or isodiametric (H. maxima) cells (Figs. 4b,d,f). The mesocarp is mostly crushed in *H. asiatica*, represented by two to three layers of parenchyma cells in H. insularis and three to five layers of elongated parenchyma cells with wavy cell walls in H. maxima. The endocarp is multi-layered, and the cells are highly lignified, but the seed coats in all the species are represented by two to three layers of degenerating cells (Figs. 4b,d,f).



PAN

Achene morphology and pericarp anatomy of Anemoninae

www.journals.pan.pl

**Fig. 4.** Cross section of *Hepatica* achenes (**a**,**b**) *H. asiatica* (**c**,**d**) *H. insularis* (**e**,**f**) *H. maxima*. Scale bars: **a**,**c**,**d** = 0.75 mm; **b**,**d**,**f** = 0.1 mm. Abbreviations: enc – endocarp, exc – exocarp, msc – mesocarp, sc – seed coat, vb – vascular bundle.



#### PULSATILLA MILL.

Four species of the genus *Pulsatilla* were observed. The results of the study indicate that *Pulsatilla* species exhibit very similar achene morphological and anatomical structures. The achenes are much longer than wider, elliptical in shape, brown to black in color, and profoundly have hirsute with white and shiny long hair (Fig. 5; Table 2). The largest measured achenes are in *P. tongkangensis*  $(4.07\pm0.24 \times 1.17\pm0.51 \text{ mm})$  and the shortest measured achenes are in *P. dahurica*  $(2.7\pm0.3 \times 0.87\pm0.08 \text{ mm})$ . The lateral wings are narrow and the persistent style forms a long slender beak.



**Fig. 5.** Achenes of *Pulsatilla* under stereomicroscope (a) *P. dahurica* (b) *P. tongkangensis* (c) *P. cernua* (d) *P. koreana.* Scale bars = 1 mm

In the cross-section, the achenes are circular in outline (Figs. 6a,c,e,g). The cross-section diameter is comparable to the width of the achenes, as in *Anemone* and *Hepatica* species (Table 3). Corresponding with the achene size, the pericarp and the endocarp are thickest in *P. tongkangensis* (58.63  $\pm$ 8.199 µm and 32.84 $\pm$ 3.35 µm, respectively),

however, the thinnest pericarp  $(32.11\pm5.78 \ \mu\text{m})$ and endocarp  $(18.24\pm3.41 \ \mu\text{m})$  are measured in *P. cernum*. The exocarp is mostly crushed or rarely present as fragmented cells in all four species (Figs. 6b,d,f,h). The mesocarp is represented by indistinguishable layers of degenerated cells, whereas the endotesta comprises a thick single layer of highly-lignified sclereid cells. The seed coat is also represented by degenerated cells, except in *P. tongkangensis* which has two- to three layers of elongated cells with thickened walls (Fig. 6d).

#### STATISTICAL ANALYSES

One-factor ANOVA was performed on nine quantitative achene traits and the differences between the species were found to be highly significant (P < 0.001) (Table 3). Pearson's correlation coefficient also indicated a significant correlation between different achene traits. The differences in the thickness of the pericarp and endocarp are presented in boxplots (Figs. 7a,b). The relationships among the species through six quantitative and 11 qualitative features were revealed using cluster analysis (Fig. 8). The cluster analysis, based on the paired group (UPGMA) algorithm using Gower similarity index, revealed three groups. Anemone narcissiflora complex represented the basal group of the UPGMA tree, three Hepatica species remained at the top of the phenograms and the third group represented by two subgroups, in which A. reflexa and A. koraiensis separated first, and A. raddeana was grouped with four Pulsatilla species.

## DISCUSSION

Molecular phylogenetic studies suggested, in a wider perception, that *Anemone* incorporated *Hepatica*, *Pulsatilla*, and *Knowltonia* (Hoot et al., 1994; Schuettpelz et al., 2002; Meyer et al., 2010). The group of these taxa also share some similar morphological characters, such as rosette basal leaves with a variety of perennating structures, an inflorescence with involucre leaves on the peduncle, petaloid sepals, and similar achene features. However, there have been differences in a few morphological features, such as close proximity to the flower and stalked achenes of *Hepatica*, an elongated plumose style in *Pulsatilla*, and berry-like fruits in *Knowltonia* and *Anemone* (Hoot et al., 1994). We compared the achene morphology and - www.czasopisma.pan.pl PAN www.journals.pan.pl \_\_\_\_\_\_ Achene morphology and pericarp anatomy of Anemoninae



**Fig. 6.** Cross section of *Pulsatilla* achenes  $(\mathbf{a}, \mathbf{b})$  *P. dahurica*  $(\mathbf{c}, \mathbf{d})$  *P. tongkangensis*  $(\mathbf{e}, \mathbf{f})$  *P. cernua*  $(\mathbf{g}, \mathbf{h})$  *P. koreana.* Scale bars:  $\mathbf{a}, \mathbf{c}, \mathbf{e}, \mathbf{g} = 0.25$  mm;  $\mathbf{b}, \mathbf{d}, \mathbf{f}, \mathbf{h} = 0.1$  mm. Abbreviations: enc – endocarp, exc – exocarp, sc – seed coat, vb – vascular bundle.



**Fig. 7.** Normal boxplot showing pericarp and endocarp thickness of achene (**a**) pericarp (**b**) endocarp.



Fig. 8. UPGMA tree based on achene features of *Anemone*, *Hepatica*, and *Pulsatilla*.

anatomy of 12 taxa belonging to three genera, excluding *Knowltonia* from *Anemone* complex.

Exomorphological comparison revealed that the achenes of the studied taxa exhibited variations in the shape, size, color, surface, wings, and the remnant of the style. The ANOVA of the quantitative variables revealed significant differences in length and width of the achene among the taxa (P < 0.001) (Table 3) and Pearson's correlation coefficients showed the positive relationship between the length and width (r=0.934, P<0.01) (Table 4). Among the qualitative features, the surface, wings, and style are highly variable within and between the genera. In previous studies, achene characters such as size, shape, and style have been proven to be useful for distinguishing the species in the genus Anemone (Saoud et al., 2007; Ziman et al., 2008, 2011; Maciejewska-Rutkowska and Antkowska, 2013) and the result of this study evidently corroborated it. In a recent study, Ghimire et al. (2020) reported that the achene size, permanent style, surface sculpture, shape in cross-section, and thickness of the exocarp and endocarp are valuable features, which can be used for species delimitation in Clematis. The five taxa of Anemone were clearly divided into two groups based on their shape, surface, and wings. Anemone narcissiflora and A. narcissiflora subsp. crinita, which belong to the sub-genus Omalocarpus, had obovoid and flattened achenes with a glabrous surface and wide wings, whereas A. koraiensis, A. raddeana, and A. reflexa, which belong to the sub-genus Anemonanthea, had elliptical achenes with a pubescent surface and narrow wings. However, the presence of a hooked style in all taxa of the Anemone differentiates the genus from Hepatica and Pulsatilla species, which had a slender style.

A similar result was obtained for achene anatomy. Achenes of Hepatica and Pulsatilla are circular in outline, whereas achenes of Anemone are oval to elliptical in cross-section. The quantitative variables indicated that the pericarp and endocarp thickness and the achene diameter in the cross--section of both planes vary significantly among the species (P < 0.001). The pericarp thickness is positively correlated with the endocarp thickness (r=0.498; P<0.01) and the diameter in the cross--section of both planes (r=0.487; P<0.01 and r=0.8; P < 0.01), whereas the endocarp thickness is negatively correlated with the achene length (r=-0.318; P < 0.01) and width (r=-0.167; P < 0.05). Among the qualitative features, the nature of the exocarp, mesocarp, and endocarp are important because the seed coat was poorly represented in all taxa.

	Length	Width	L/W	Pericarp	Endocarp	P/E	CSD1	CSD2	D1/D2
Length									
Width	.934**								
L/W	334**	605**							
Pericarp	095	.062	411**						
Endocarp	318**	167*	308**	.498**					
P/E	007	.042	185**	.510**	317**				
CSD1	.674**	.838**	820**	.487**	.229	.098			
CSD2	.050	.259*	699**	.800**	.765**	.077	.662**		
D1/D2	.852**	.875**	492**	025	333**	.063	.766**	.042	

TABLE 4. Pearson's correlation coefficients between different seed features in Anemoninae species. L/W = length and width ratio, P/E = pericarp and endocarp ratio, CSD1 = diameter parallel to cotyledon, CSD2 = diameter perpendicular to cotyledon, D1/D2 = CSD1 and CSD2 ratio.

\*\*Sig. at 0.01 level \*Sig. at 0.05 level

In cross-section outline, Anemone achenes are oval or elliptical with narrow to wide extensions wherein the vascular bundles are present. The achene anatomy of Anemone was extensively studied in the past. Ziman et al. (2008) and Maciejewska-Rutkowska and Antkowska (2013) emphasized the taxonomic value of the pericarp in this genus. Before that, Chaudhary and Trifonova (1988) distinguished the epicarp, mesocarp, and endocarp of 11 species of Anemone and highlighted the significance of the number of cell layers and cell forms in individual pericarp layers. Even with a very limited sampling (five taxa), we found a similar result in this study. The exocarp is cutinized with rectangular to elongated cells in A. koraiensis and A. reflexa, but the cells are crushed in the other three taxa; however, a few fragmented exotestal cells were found in A. narcissiflora. A similar trend was observed for the mesocarp, except for the number of cell layers. However, the most characteristic feature in Anemone seems to be the nature of the endocarp, which clearly separated the two subgenera in our sampling. The endocarp is exclusively single-layered and the cells are highly lignified and palisade-like in A. raddeana and A. reflexa, which belong to the subgenus Anemonanthea. However, cells are highly lignified, sclereid and single-layered in A. narcissiflora, and two- to three-layered in A. narcissiflora subsp. crinita, which belongs to the subgenus Omalocarpus. The endocarp is a thin layer of thick-walled rectangular cells in A. koraiensis. According to Ghimire et al. (2020), the endocarp is the most consistent region in the Clematis species. Morphologically, A. koraiensis is closely related to

*A. reflexa* and both species share similar vegetative forms, a white solitary flower with five sepals and a pubescent achene surface (Sung, 2017). The results of this study agree with those of Sung (2017) because the two species formed a distinct subclade on the *Pulsatilla-Anemone* clade in the UPGMA phenogram.

In Hepatica, the exotesta is well characterized by tangentially-elongated cells, as in H. asiatica and H. insularis, or by broadly rectangular cells, as in H. maxima. Following the exocarp, H. maxima also revealed three to five cell layers-thick mesocarp and a maximum of four-cell layered endocarp, however, H. asiatica and H. insularis exhibited one to two layered mesocarp and a maximum of three-layered endocarp. The result of this study supported the classification of Nakai (1952), who divided these three Korean species into two groups: H. asiatica and H. insularis with annual leaves and H. maxima with biennial leaves. In addition, Pfosser et al. (2011) found a genetically close relationship between H. asiatica and H. insularis. The qualitative achene features of four Pulsatilla species included in this study are astonishingly similar in every detail. It is not surprising because in a recent taxonomic study by Grey-Wilson (2014), all four investigated species belonged to the same section Semicampanaria of the subgenus Pulsatilla.

In conclusion, regardless of limited species samples, our results suggested that achene morphology and anatomy provide a useful taxonomic base for species delimitation in the studied genera. *Anemone* species showed a remarkable variation in the nature of the endotesta that could contribute to the subgeneric classification of the genus. In addition, the pericarp structure, particularly the number of cell layers and cell forms in the exocarp and endocarp, could be used as an alternative source of species discrimination in *Anemome* and *Hepatica*.

# AUTHORS' CONTRIBUTIONS

Conceptualization: Balkrishna Ghimire and Mi Jin Jeong; Experiments: Balkrishna Ghimire; Statistical analysis: Dabin Yum and Balkrishna Ghimire,; Manuscript writing: Balkrishna Ghimire; Review and editing: Dabin Yum, Jae Hyun Kim and Mi Jin Jeong; Funding acquisition: Jae Hyun Kim and Mi Jin Jeong.

# ACKNOWLEDGEMENTS

This study was financially supported by the project 'Ex-situ Conservation of Forest Plant Seeds in Korea (KNA1-2-29).

# REFERENCES

- AMINI E, ZARRE S, and ASSADI M. 2011. Seed micro--morphology and its systematic significance in *Gypsophila* (Caryophyllaceae) and allied genera. *Nordic Journal of Botany* 29: 660–669.
- APG IV. 2016. An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG III. *Botanical Journal of the Linnean Society* 181: 1–20.
- CAPPELLETTI EM, and POLDINI L. 1984. Seed morphology in some European aconites (Aconitum, Ranunculaceae). Plant Systematics and Evolution 145: 193–201.
- CHAUDHARY RP, and TRIFONOVA VI. 1988. Morphology of fruit and comparative anatomy of pericarp and seed coat in the Nepal species of the genus *Anemone* (Ranunculaceae). *Botanicheskii Zhurnal* 73(6): 803–817.
- EMADZADE K, LEHNEBACH C, LOCKHART P, and HÖRANDL E. 2010 A molecular phylogeny, morphology and classification of genera of Ranunculeae (Ranunculaceae). *Taxon* 59: 809–828.
- FRANCHET A. 1889. *Plantae delavayae*, Vol 1. Paul Klineksieck, Paris, pp 6–7.
- FUKUHARA T. 1999. Seed and funicle morphology of Fumariaceae-Fumarioideae: systematic implications and evolutionary patterns. *International Journal of Plant Scince* 160: 151–180.

- GHIMIRE B, JEONG MJ, CHOI GE, LEE H, SUH GK, HEO K, and KU JJ. 2015. Seed morphology of the subfamily Helleboroideae (Ranunculaceae) and its systematic implication. *Flora* 216: 6–25.
- GHIMIRE B, JEONG MJ, LEE KM, HEO K, LEE H, and SUH GK. 2016a. Achene morphology of Saussurea species (Asteraceae, Cardueae) in Korea and its systematic implications. Botanical Journal of the Linnean Society 181: 692–710.
- GHIMIRE B, LEE H, CHOI GE, JEONG MJ, LEE CH, SUH GU, HEO K, and SON SW. 2016b. Seed morphology of 12 taxa of the genus *Thalictrum* L. (Thalictroideae, Ranunculaceae) and its systematic implication. *Phytotaxa* 283: 271–285.
- GHIMIRE B, PARK BK, SON DC, and OH SH. 2020. Achene morphology and anatomy of *Clematis* L. (Ranunculaceae) in Korea and its taxonomic implications. *Plants* 9: 1279.
- GREY-WILSON C. 2014. Pasque-flowers. The genus Pulsatilla. Kenning hall: The Charlotte Louise Press.
- HADIDCHI A, ATTAR F, and ULLAH F. 2019. Using microscopic techniques for taxonomic implications of seed and fruits of *Delphinium* L. (sensu lato) Ranunculaceae. *Microscopy Research and Technique* 83(2): 1–19.
- HAMMER Ø, HARPER DAT, and RYAN PD. 2001. PAST: Paleontological Statistics Software Package for Education and Data Analysis. *Palaeontologia Electronica* 4(1): 9pp.
- HEISS AG, KROPF M, SONTAG S, and WEBER A. 2011. Seed morphology of *Nigella* s.l. (Ranunculaceae): Identification, diagnostic traits, and their potential phylogenetic relevance. *International Journal of Plant Science* 172: 267–284.
- HEYWOOD VH, BRUMMITT RK, CULHAM A, and SEBERG O. 2007. Plant families of the world. Richmond Hill, Ontario, Canada: Firefly Books.
- HOOT SB, REZNICEK AA, and PALMER JD. 1994. Phylogenetic relationships in *Anemone* (Ranunculaceae) based on morphology and chloroplast DNA. *Systematic Botany* 19: 169–200.
- HUTCHINSON J. 1923. Contributions towards a phylogenetic classification of flowering plants Bulletin of Miscellaneous Information Royal Botanic Gardens Kew 2, 65–89.
- JANCHEN E. 1949. Die systematische Gliederung der Ranunculaceen und Berberidaceen. Denkschriften der Österreichischen Akademie der Wissenschaften 108(4): 1–82.
- JENSEN U, HOOT SB, JOHANSSON JT, and KOSUGE K. 1995. Systematics and phylogeny of the Ranunculaceae – a revised family concept on the basis of molecular data. *Plant Systematics and Evolution* (Suppl.) 9: 273–280.
- JIANG N, ZHOU Z, YANG JB, ZHANG SD, GUAN KY, TAN YH, and YU WB. 2017. Phylogenetic reassessment of tribe Anemoneae (Ranunculaceae): non-monophyly of Anemone s.l. revealed by plastid datasets. PLoS One 12, e0174792. doi: 10.1371/ journal. pone. 0174792

- JOHANSSON JT and JANSEN RK. 1993. Chloroplast DNA variation and phylogeny of the Ranunculaceae. *Plant Systematics and Evolution* 187: 29–49.
- JUAN R, PASTOR J, and FERNANDEZ I. 2000. SEM and light microscope observations on fruit and seeds in Scrophulariaceae from Southwest Spain and their systematic significance. *Annals of Botany* 86: 323– 338.
- JUNG WC, and HEO K. 2017. Fruit and seed morphology of Korean Ranunculaceae. Korean Journal of Plant Taxonomy 47: 137–153.
- LEHTONEN S, CHRISTENHUSZ MJ, and FALCK D. 2016. Sensitive phylogenetics of *Clematis* and its position in Ranunculaceae. *Botanical Journal of the Linnean Society* 182: 825–867. doi: 10.1111/b0j.12477
- MACIEJEWSKA-RUTKOWSKA I, and ANTKOWSKA W. 2013. Taxonomic utility of achene morphology and anatomy in *Anemone* L. (Ranunculaceae) species. *Acta Biologica Carcoviensia Series Botanica* 55: 29–36.
- MEYER KM, HOOT SB, and ARROYO MTK. 2010. Phylogenetic affinities of south American Anemone (Ranunculaceae), including the endemic segregate genera, Barneoudia and Oreithales. International Journal of Plant Science 171: 323–331. doi: 10.1086/650153.
- NAKAI T. 1952. A synoptical sketch of Korean flora. Bulletin of Natural Science Museum 1–27.
- PFOSSER M, SUN B-Y, STUESSY TF, JANG C-G, GUO Y-P, TAEJIN K, HWAN KC, KATO H, and SUGAWARA T. 2011. Phylogeny of *Hepatica* (Ranunculaceae) and origin of *Hepatica* maxima Nakai endemic to Ullung Island, Korea. Stapfia 95:16–27.
- REN Y, LI ZJ, CHANG HI, LEI YJ, and LU AM. 2004. Floral development of Kingdonia (Ranunculaceae s. l., Ranunculales). Plant Systematics and Evolution 247:145–53.
- SAOUD SN, KNIO K, and JURY S. 2007. Phenetic analysis of Anemone coronaria (Ranunculaceae) and related species. Botanical Journal of the Linnean Society 153: 417–438.
- SCHUETTPELZ EJ, HOOT SB, SAMUEL R, and EHRENDORFER F. 2002. Multiple origins of Southern Hemisphere Anemone (Ranunculaceae) based on plastid and nuclear sequences. Plant Systematics and Evolution 231: 143–151.
- SIMPSON MG. 2006. Plant systematics. Elsevier, Amsterdam.

- SOLTIS DE, SOLTIS PS, ENDRESS PK, and CHASE MW. 2005. *Phylogeny and evolution of angiosperms*. Sinauer Associates, Sunderland, Massachusetts.
- STARODUBTSEV VN. 1991. Anemone: Systematics and evolution. Leningard: Akad. Nauk SSSR
- TAMURA M. 1995. Angiospermae: Ordnung Ranunculales. Fam. Ranunculaceae In: Hiepko P [ed.], *Die Natürlichen Pflanzenfamilien*. Vol. 17a(4), 324–349. Duncker & Humblot, Berlin.
- SUNG HY. 2017. Anemone L. Flora of Korea: Volume 2a, Magnoliidae, p. 60-64.
- THE PLANT LIST. 2013. Version 1.1. Published on the Internet; http://www.theplantlist.org/ (accessed 1st January).
- WANG WT. 1964. Duo genera nova Rannuculacearum Sinensium. Acta Phytotaxon Sinica 9: 103–7.
- WANG WT. 1979. Tribe Anemoneae DC. In: Anonymous (ed) Flora Reipublicae Popularis Sinica, Vol 28. Science Press, Beijing, pp 1–141.
- WANG W, LU A-M, REN Y, ENDRESS ME, and CHEN Z-D. 2009. Phylogeny and classification of Ranunculales: Evidence from four molecular loci and morphological data. Perspectives in Plant Ecology, Evolution and Systematics 11: 81–110.
- Xu F. 2003. Sclerotesta Morphology and its systematic implications in Magnoliaceous seeds. *Botanical Journal of the Linnean Society*. 142: 407–424.
- YANG QE. 2002. Cytology of ten species in Anemone, one in Amemoclema and six Clematis (Trib. Anemoneae, Ranunculaceae) from China. Acta Phytotaxonomica Sinca 40: 396–405.
- ZHANG ZY, YANG DZ, Lu AM, and KNAPP S. 2005. Seed morphology of the tribe Hyoscyameae (Solanaceae). *Taxon* 54: 71–83.
- ZIMAN SN, BULAKH EV, KADOTA Y, and KEENER CS. 2008. Modern view on the taxonomy of the genus Anemone L. sensu stricto (Ranunculaceae). Journal of Japanese Botany 83: 127–155.
- ZHANG GL, and GONG X. 2002. The karyotype analysis of Anemoclema glaucifolium and Heteroplexis microcephala both endemic to China. Acta Botanica Yunnanica 24: 765–768.
- ZHANG Y, KONG HH, and YANG QE. 2015. Phylogenetic relationships and taxonomic status of the monotypic Chinese genus *Anemoclema* (Ranunculaceae). *Plant Systematics and Evolution* 301: 335–44.